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A REVIEW ON THE POTENTIALITY OF ANTI-MICROBIAL AND ANTI-CANCER COMPOUNDS PRODUCED BY MYXOBACTERIAL SPECIES.

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ABSTRACT

Myxobacteria, a get-together of Gram-negative aerobes, and are people from d-Proteobacteria and are encapsulated by tremendous genomes, efficient social lead, coasting motility, and starvation-provoked fruiting body advancement. Natural products created by myxobacteria display a tremendous basic assorted variety with observable organic exercises. The myxobacterial compound, Epothilone, it adjust microtubules inside the cell, with the objective that utilitarian mitotic shafts can never again be amassed. As Epothilone moreover kills multidrug safe cells, and is well dissolvable in water, and can be made by maturing, it is a promising contender for an antitumor medicine. In the latest decades, a regularly expanding number of research focuses have shown extraordinary excitement for myxobacteria metabolites, especially epothilones. Epothilones are starting at now in arranging III clinical fundamentals as anticancer experts, while isoxazole and the tubulin are in preclinical improvement.

1. INTRODUCTION

Myxobacteria are gram negative, single-celled rod shaped bacteria which are found inhabiting various environmental niches like soil, the rotting plant

mass and the outer layer of the stem and root of the wood plant which. Myxobacteria are important part of soil microbes and are members of the delta-proteobacteria family (Dawid, 2000). Myxobacteria are famous for their gliding behavior and potential to form fruiting bodies majorly during stress conditions. Compared to other bacteria, myxobacteria have some detectable features: The ability to glide on solid surface, swarming motility, degrading macromolecules, preying on living microorganisms. Myxobacteria are considered to be social bacterial species, as they form herds and mass hunt for other microorganisms. Myxobacteria are multicellular, enabling a quality particularly useful for the study of signaling between cells (Goldman *et.al.*, 2006). There is an industrial interest for the production of secondary metabolites from myxobacterial species (Weismann and Muller 2010). Myxobacterial species are majorly found in soil, where they can populate the manure of herbivorous plant materials, but metagenomic data together with actual isolates identified the alongshore and marine habitats as others, yet unaccountable habitats. Most popular study typical of this order is *Myxococcus Xanthus* DK1622 which is a representative organism for conducting various studies on this kingdom of bacteria. Numerous Myxobacterial species produce lytic catalytic enzymes which enable them to feed on other microscopic organisms and to degrade the proteins, and nucleic acids. Myxobacterial species show swarming movements for acquiring food and due to the non-appearance of any supplement, myxobacteria end the swarming conduct and begin to develop bunches, ultimately resulting in the formation of fruiting bodies. Complex life cycle that incorporates the arrangement of Myxospores inside the grown-up fruiting bodies is one of the brilliant highlights of this species. Myxobacterial species belonging to genus *Myxococcales* are further divided into suborders including *cystobacterineae*, *nanocystineae*, and *sorangineae*. Myxobacterial species form fruiting bodies that show brilliant hues and can be viewed on the media plates. The principal mycobacterium, *Polyangium vitellinum*, was found and named by the German botanist H.F. Connection in 1809 (Link, H.F. 1809; Wolfgang Dawid.2000). The American botanist Roland Thaxter, in 1892, was the first to distinguish these species as myxobacteria and portray their life cycles (Thaxter, R. 1892; Wolfgang Dawid.2000). The mind boggling life cycle that includes the arrangement of Myxospores inside the develop fruiting body is one of the exceptional highlights of myxobacteria. Another extraordinary element of myxobacteria which may be the explanation of its perplexing life cycle when contrasted with other microscopic organisms is the general enormous size of myxobacterial genomes (9500–10,000 kbp) and a DNA with a G&C substance of 66–72%. The sequenced genome of *Sorangium Cellulosum* Soce56 looks like the so far biggest known bacterial genome with a size of 13.0338 Mb. The uncommon enormous estimate just as the high number of qualities in the *Myxococcus* genome in contrast with all other so far sequenced non myxobacterial δ Proteobacteria should be at any rate to some degree dependent on broad quality duplications. These quality duplications don't advance unintentionally yet rather incorporate qualities which are significant for the myxobacterial life cycle, for example, qualities encoding proteins important for intercellular flagging.

Myxobacteria integrate countless organically dynamic auxiliary metabolites. A significant number of those mixes were new. Most myxobacterial substances are tolerably lipophilic, direct or cyclic polypeptides' and peptides. The peptides regularly are depsipeptides containing hydroxyl acids notwithstanding now and then extremely strange amino acids: e.g., amino acids, 4-methylazetidincarboxylic corrosive, homoproline, and much increasingly intriguing examples. The myxobacterial compound, Epothilon, it balances out microtubuli inside the cell, with the goal that utilitarian mitotic shafts can never again be assembled. With mitosis made unimaginable, the cell enters apoptosis. As Epothilon additionally murders multidrug safe cells, and is well solvent in water, and can be created by aging, it is a promising contender for an antitumor medication. Another new compound, tubulysin, has the contrary impact on tubulin: it prompts a total breakdown of the microtubuli inside hours (Sasse, Khalil et al., GBF, and unpublished information). One myxobacterial substance, soraphen, even has a novel instrument of activity. This substance squares explicitly and with high adequacy parasitic acetyl - CoA carboxylase.

1. 2. Scientific Classification:

KINGDOM	PHYLUM	CLASS	ORDER	SUBORDER	FAMILY	GENUS	SPECIES
<i>Bacteria</i>	<i>Proteobacteria</i>	<i>Proteobacteria</i>	<i>Myxococcales</i>	<i>Cystobactereae</i>	<i>Myxococcaceae</i>	<i>Pyxicoccus</i> <i>Corallococcus</i> <i>Myxococcus</i>	<i>Xanthus</i>
					<i>Cystobactereae</i>	<i>Melittangium</i> <i>Stigmatella</i> <i>Cystobacter</i> <i>Hyalangium</i> <i>Archangium</i>	
				<i>Sorangineae</i>	<i>Polyangiaceae</i>	<i>Sorangium</i> <i>Polyangium</i> <i>Byssophage</i> <i>Haploangium</i> <i>Chondromyces</i>	

						<i>Jahnia</i>
				<i>Nanocystin eae</i>	<i>Nannocyst aceae</i>	<i>Nannocysti s Kofleria Haliangiu m</i>
					<i>Kofleriace ae</i>	

3. GENOME AND PHYLOGENY

The Phylogenetic tree of myxobacteria is dependent on examination of 16 rRNA quality arrangements of 101 strains that speak to six families, 3 subfamilies, twenty genera, forty six species and twelve other novel taxa. It has long been clear that these myxobacteria are divided into 2 groups, which are distinguished from each other by morphological and physiological parameters. The genome of myxobacteria is normal microscopic ring chromosome, and contains about twice upwards of 9450 kbp about the estimate and state of *Escherichia coli*, Genus *streptomycin*. Myxobacterial DNA has a GC content somewhere in the range of 64 and 72 mol%; The GC content in the substance in the suborder *sorangineae* is marginally higher than that of *cystobacterineae* (64-70 mol %). The GC content fluctuates significantly between various DNA fragment for example in *M.Xanthus* among 36 and esteems more than 80 % (Komano, T.et al, 1987) Fig shows the total quality guide of the genome of *M.Xanthus* strain DK1622.

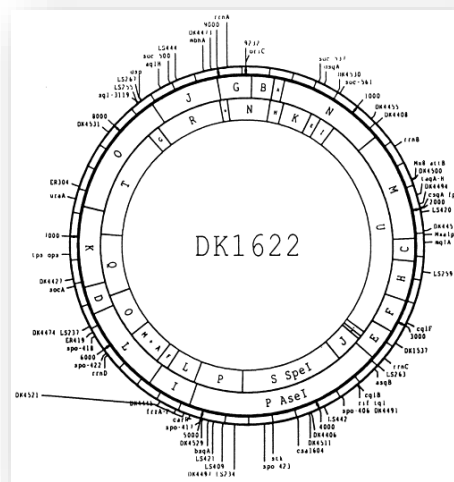


Fig 1: Physical and hereditary guide of *M.Xanthus* (strain DK1612) chromosome (Reichenbach, 1999).

Myxobacterial species *M.Xanthus* and *Stigmatella Aurantiaca* were the principal prokaryotes where another class of retro element, called retrons, has found. Encodes turn around transcriptase practically equivalent to those

emerging in retroviruses. They likewise code for surprising satellite DNA created by invert interpretation as multicopy single defamed DNA. Their basic event with sulfated and sulfur-diminishing microscopic organisms and with bdellovibrions in the delta part of Proteobacteria might be conjectured concerning whether myxobacteria speak to a vigorous relative of sulfate or sulfur-lessening microorganisms. Succession contrasts between the 5s and 16s rRNA particles propose that the delta branch may have started 750 to 1100 million years prior (Darwin, M. 1996).

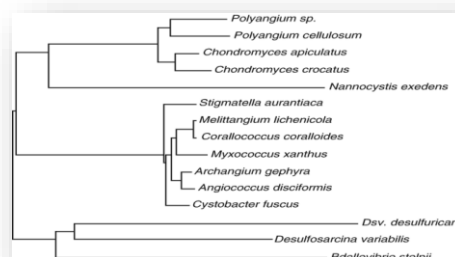


Fig 2: Phylogenetic tree of the myxobacteria. The tree has been gotten from evolutionary distances of the 16s rRNA particles (Shimkets and Woese, 1992).

4. Isolation and Cultivation of Myxobacteria

Myxobacterial species have been isolated from soil samples and rotting plant samples. The most important technique used for the isolation is the *E.coli* bedeviling strategy (Gaspari et al., 2005). Environmental samples (e.g. sand, soil, dead wood, manure) are usually used for the isolation of these bacterial species. The isolation and development of potential and fascinating new gainful strains requires the utilization of another technique alongside essential routine strategies. In this way some exceptional procedure to isolate myxobacteria from soil, for example, baiting with excrement pellets (Reichenbach and Dworkin, 1992), brooding of bark in the moist chamber (Peterson 1969), baiting with *E.coli* streaks (Reichenbach and Dworkin), immunization of channel paper with soil (Reichenbach and Dworkin 1992). Isolation and cultivation is difficult, several kinds of media such as PDCY Media, V/Y Media have been tried but better results were given by CYE Media. The medium comprises of 0.1% Casitone (Difco Laboratories, Detroit, MI, USA), 0.03% yeast extract, 1.5% agar, 1.5% NaCl and half quality SWS solution (Iizuka et al., 1998).

Table 1: Overview of Myxobacterial species found in soils Table legend is always over the table.

GENUS	SPECIES
<i>Myxococcus</i>	<i>Fulvus, stipitatus, virescens, xanthus</i>
<i>Sorangium</i>	<i>Aurantiaca, erecta</i>
<i>Archangium</i>	<i>Gephyra, serpens</i>
<i>Corallococcus</i>	<i>Coralloides, exiguus</i>
<i>Cystobacter</i>	<i>Ferrugineus, fuscus, minus, velatus</i>
<i>Haploangium</i>	<i>Spp.</i>
<i>Nannocystis</i>	<i>exedens</i>

5. FORMATIONS OF FRUITING BODIES AND MYXOSPORES

Myxobacterium has amazing features is that form the fruiting body under famine conditions. Starvation triggered the development of the body, and the structure is completely formed by cell movement. The cooperative morphology of hundreds or thousands of cells first leads to their aggregation and formation of large mounds and eventually yield called fruiting bodies. They have simple appearance such as Myxococcus, the fruiting body is mushy slimy consistency and spherical. The shade of fruiting bodies possibly orange, white, red, darker, and so forth the shape and size, shading and course of action of fruiting bodies are utilized as a trait of explicit, ordinary assurance. The advancement of fruiting bodies is driven by dietary structure and is constrained by pH, supplement fixation, confines, and temperatures. Vegetative myxobacteria cells inside fruiting bodies transform into small, alternatively refractory Myxospores, which have been able to tolerate adverse condition for decades. It is a cooperative form by the flock cells of the plant. The process is a sequence of the following steps:

- I) During morphogenesis, countless swarming cells slacken their physiological character.
- II) Vegetative development of the rod closes.
- III) The cell in some situations of the swarm accumulates to frame totals.
- IV) Molecules that structure on the outside of the cell, cells remain together.
- V) The outcome is an unstructured agglutination of the mass of cells that is unconstrained to roughly 75-90%.

- VI) Construction of exceptional auxiliary component (stem, base plate, sporangial divider) starts.
- VII) An attribute of a fruiting body is shaped.
- VIII) During the development stage, vegetative bar cells change into Myxospores by cell morphology.

The whole procedure of fruiting body development takes 12-14 hours under ideal conditions. *S.aurantiaca* is considered as a model for photo morphogenesis in myxobacteria shaping bodies: the structure of fruiting bodies and the somewhat reliant development cycle have been seriously contemplated. Aside from ecological components, the outside flagging particles, pheromone, decides the improvement cycle. These are low atomic weight lipophilic mixes radiated by *Sg. Aurantiaca* on the impact of a supplement does subterranean insects their development is incredibly upgraded by light. Pheromones have body stimulating activities. Myxospores are formed inside maturing sporangioles and fruiting bodies. Vegetative rod cell undergo cellular morphology small and round. Whole vegetation the cells transform into a Myxospores.

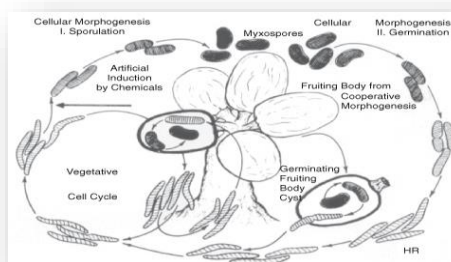


Fig3: Cellular morphogenesis of myxobacteria, appeared by the case of *S. aurantiaca* (Wireman and Dworkin, 1975).

The adjustment fit as a fiddle is self-evident: Myxospores are a lot littler and thicker than vegetative cells, they show up round, emphatically light-refracting and encompassed by a meager case. Antibacterial endospores from Myxospores contrast as endurance cells (for example Class Bacillus) by structure, physical properties, and development strategy. The reason for Myxospores is the presence of myxobacteria during times of unfavorable remains, for example, cold and summer period, dryness, corrosive pH or anoxic conditions. Suspension in phosphate cradle Myxospores can be brought to grow, not to solidify the body. The motivation behind Myxospores is the presence of myxobacteria during the time of antagonistic remains, for example, cold and summer period, dryness, corrosive pH or anoxic conditions. The motivation behind fruiting bodies is to begin another life cycle with a bigger populace, to at the same time hydrolyze biosimilar utilizing an exorma chemical, and to utilize such wholesome sources with the most extreme association.

6 Secondary Metabolites

Unique item against organisms likewise assumes a significant job in the identification and advancement of human irresistible illness medicate plan (Cragg and Newman, 2013). Microorganisms catalyze fundamental metabolites, for example, sugars, amino acids, nucleotides that demonstration exceptionally significant for their progression and development. Secondary metabolites are usually generated at the level of late development by fixed stresses related to microorganism species. They produce them for their own welfare, which they used to kill their prey in order to make symbiosis with plant and animals to remain in adverse condition or to kill their prey or to save them from injury. The principal makers of secondary metabolites among microbes incorporate actinomycetes, bacillus, and pseudomonades (Garth *et.al.*, 2003). Myxobacteria is viewed as a rich wellspring of fundamentally assorted optional metabolites experiencing intriguing organic activity (Herrmann *et.al.*, 2017). Almost all bioactive secondary metabolites produced by myxobacteria have antibacterial, antifungal, anticancer activity (Kim *et.al.*, 2003).

All myxobacteria are depicted by their capacity to debase natural macromolecules. With respect to the utilization of cellulose, they structure two gatherings dependent on their capacity to utilize inorganic nitrogen mixes. The bactericidal capability of myxobacteria is produced during development and seclusion from the soil likewise from the compost of plant-eating creatures. In spite of their cellulolytic alongside batriolytic activity, myxobacteria are receptive to anti-microbial in being delicate to erythromycin, neomycin, kanamycin, streptomycin, and antibiotic medication. Around all myxobacteria are touchy to actinomycin, marginally remarkable for gram-negative microbes. Around 80% of all naturally dynamic auxiliary metabolites are incorporated by prokaryotes (for example Streptomycin, bacillus, pseudomonas species), about 20% by eukaryotes (for the most part parasites). By and by there is somewhere in the range of 10000 bioactive compounds of microorganism cause.

Myxobacteria produce an enormous number of such bioactive particles with antifungal, synthetic just as antitumor activity. More than eighty infrastructures have been identified so far with around 350 structural adaptations, some of which may have medical application potential. The biosynthetic limit of myxobacteria is somewhat chefsul: 50-100% of all myxobacteria produce a compound with natural exercises, having a place with polypeptides from different nonionic classes, for example, acetic acid derivation and propionate, straight and cyclic peptide, a heterocyclic compound, and so forth. Twenty years back the primary synthetic structure of Myxobacterial anti-toxin ambrucitin-shaped by *Mx. fulvovus* was known as myxothiazol. Likewise 20 basic variations' of myxovirescin framed by *M.virescens* and more than 50 soraphen shaped by *So. Cellulosum* was investigating .Following is the some example for the activities of bioactive substance from myxobacteria:

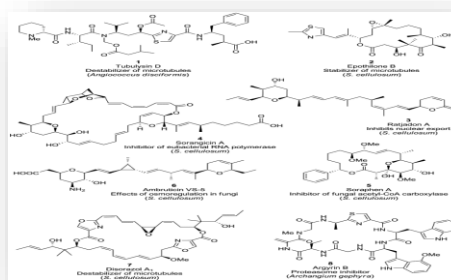


Fig4: Synthetic structure dependent on some bioactive optional metabolites of myxobacteria (Weismann and Müller, 2010).

Three artificially particular new substances and a basic theme have been shrouded that square RNA polymerase: myxosporin, a strain of *Mx fulvovus* in 1983 and corollapirin, a strain of *CC.coralloides*, which is regularly shaped by the strain of *CC.coralloid*. Both follow up on gram-positive and don't infiltrate gram-negative cells. About the three myxobacteria anti-microbial that demoralize protein combination in eukaryotes, myxowalargin has been the best examined. It was found in a strain of *Mx.fullview*, yet additionally in a strain of *Mx.fullview*, yet in addition in a strain of *CC.coralloides* and *Ar.gephyra*. It kills yeast other organisms just as has anti-toxin activity against gram-positive and gram-negative microbes. Myxothiazol by a strain of *Mx fulvovus* hinders the cytochrome complex in the respiratory chain, while NADH ubiquinone oxidoreductase if repressed by myxalamide (1983) from *M.Xanthus* and *S.aurantiaca* from aurachine (1987). These substances have antifungal exercises. So in 1993 sorafen separates from *S.cellulosum*: it squares acetyl-CoA carboxylase, the chloroplast catalyst accepting on phytopathogenic organisms. In 1987 a promising stuff was confined from *S.cellulosum*: Epothilones. It follows up on the cytoskeleton of the eukaryotic cells, squares cell division and prompts cell demise. Its development range is restricted: microscopic organisms are not harassed. Epothilones follows up on disease cells. It hindered the development of numerous human malignancy cell lines, for example, bosom, digestive system, and ovarian disease. There is a decent possibility that Epothilon will get signed for the treatment of disease in clinical application. In conclusion, it ought to be expressed that the shading of myxobacteria (violet, darker, red) that gather solely in the fruiting body are auxiliary metabolites.

7 Secondary Metabolites with Anticancer Activity

Characteristic items still assume a significant job in the significant revelation and advancement procedure of medications against human sicknesses especially in the field of hostile to infective and anticancer research. Myxobacteria are known to be a rich wellspring of basically various secondary metabolites (Herrmann *et.al.*, 2017). The greater part of the bioactive secondary metabolites delivered by myxobacteria has antibacterial, antifungal, anticancer exercises (Kim *et al.*, 2003). On account of their capacity to create such explicit different metabolites, they

are likewise called 'microbial processing plants' (Reichenbach, 2001). In the most recent decades, an ever-increasing number of research centers have demonstrated incredible enthusiasm for myxobacteria metabolites, particularly epothilones.

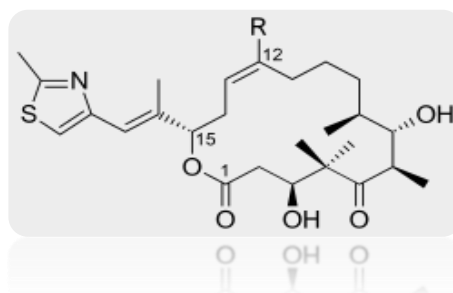


Fig5: Chemical structure of Epothilone (Reichenbach and Höfle, 1993).

The epothilones, isoxazole, chondramide B, apicularen, gephyronic corrosive, and rhizopodin are bioactive characteristic items segregated from myxobacteria. Epothilones are as of now in stage III clinical preliminaries as anticancer specialists, while isoxazole and the tubulin are in preclinical improvement.

TABLE 2: Important compound & their biological activity found in myxobacteria (Wiebke Landwehr *et. al.*).

Compound	Species	Activity	Mode of action	Reference
Epothilones	<i>S.cellulosum</i>	cytotoxic	Inhibition of microtubule	Gerth <i>et.al.</i> ,(1996)
Melithiazols	<i>M.stipitatus</i> <i>M.lichenicola</i>	antibacterial	Inhibit NADH oxidation	Sasse <i>et.al.</i> ,(1999)
Ambrucitin	<i>S.cellulosum</i>	antifungal	Interfere with high osmolarity glycerol signaling pathway	Rangel <i>et.al.</i> ,(1997) Connor <i>et.al.</i> ,(1977)
Aurachins	<i>S.aurantiaca</i>	antibacterial	Block NADH oxidation	Kunze <i>et.al.</i> ,(1987)
Crocacin	<i>C.crocatus</i>	antibacterial	Inhibit electron transport	Kunze <i>et.al.</i> ,(2014)
Etnangien	<i>S.cellulosum</i>	antibacterial	Inhibit nucleic acid polymerase	Irschik <i>et.al.</i> ,2007)
Ripostatin	<i>S.cellulosum</i>	antibacterial	Inhibit RNA polymerases	Irschik <i>et.al.</i> , (1995)
Myxothioazol	<i>M.fulves</i>	antifungal	Inhibit electron transport	Gerth <i>et.al.</i> ,(1980)

Melithiazols	<i>M.lichenicola</i> , <i>A.gephyra</i> <i>M.stipitatus</i>	antibacterial	Inhibit NADH oxidation	Sasse <i>et.al.</i> ,(1999)
Rhizopodin	<i>M. stipitatus</i>	Cytostatic	Alteration of protein phasphorilation	Sasse <i>et.al.</i> ,(1993)
Sorangicin	<i>S.cellulosum</i>	Antibacterial	Inhibits RNA polymerase	Irschik <i>et.al.</i> , (1995)
Cystobactamids	<i>C. fuscus</i>	Antifungal/cytotoxic	Inhibit submitochondrial NADH oxidation	Ojika <i>et.al.</i> ,(1998)

8 Conclusions and Future Work

Numerous parts of myxobacterial science have no other known parallels in the bacterial world and give a developmentally antiquated answer for some worldwide formative and social issues. As we proceed towards the centennial of the revelation of the myxobacteria, unmistakably despite everything we have a lot to find out about these interesting living things and Concluded that a point by point investigation of myxobacteria might be productive in finding new anti-infection agents.

References

- [1] Dworkin, M. (2000). Introduction to the myxobacteria. In *Prokaryotic Development* (pp. 221-242). American Society of Microbiology.
- [2] Dworkin, M. (2001). Myxobacteria. *e LS*.
- [3] Dworkin, M. A. R. T. I. N. (1966). Biology of the myxobacteria. *Annual Reviews in Microbiology*, 20(1), 75-106.
- [4] Fluegel, W. A. L. T. E. R. (1964). Induced fruiting in myxobacteria. In *Proc. Minn. Acad. Sci* (Vol. 31, pp. 114-115).
- [5] Garcia, R., Pistorius, D., Stadler, M., & Müller, R. (2011). Fatty acid-related phylogeny of myxobacteria as an approach to discover polyunsaturated omega-3/6 fatty acids. *Journal of bacteriology*, 193(8), 1930-1942.
- [6] Gerth, K., & Müller, R. (2005). Moderately thermophilic Myxobacteria: novel potential for the production of natural products isolation and characterization. *Environmental microbiology*, 7(6), 874-880.
- [7] Jiang, D. M., Wu, Z. H., Zhao, J. Y., & Li, Y. Z. (2007). Fruiting and non-fruited myxobacteria: a phylogenetic perspective of cultured and uncultured members of this group. *Molecular phylogenetics and evolution*, 44(2), 545-552.
- [8] Kaiser, D., Manoil, C., & Dworkin, M. (1979). Myxobacteria: cell interactions, genetics, and development. *Annual Reviews in Microbiology*, 33(1), 595-639.
- [9] Kunze, B., Böhlendorf, B., Reichenbach, H., & Höfle, G. (2008). Pedein A and B: production, isolation, structure elucidation and biological properties

- of new antifungal cyclopeptides from *Chondromyces pediculatus* (Myxobacteria). *The Journal of antibiotics*, 61(1), 18.
- [10] Livingstone, P. G., Morphew, R. M., & Whitworth, D. E. (2017). Myxobacteria are able to prey broadly upon clinically-relevant pathogens, exhibiting a prey range which cannot be explained by phylogeny. *Frontiers in microbiology*, 8, 1593.
- [11] Pradella, S., Hans, A., Spröer, C., Reichenbach, H., Gerth, K., & Beyer, S. (2002). Characterisation, genome size and genetic manipulation of the myxobacterium *Sorangium cellulosum* So ce56. *Archives of microbiology*, 178(6), 484-492.
- [12] Qualls, G. T., Stephens, K., & White, D. (1978). Morphogenetic movements and multicellular development in the fruiting myxobacterium, *Stigmatella aurantiaca*. *Developmental biology*, 66(1), 270-274.
- [13] Höfle, G., Glaser, N., Kiffe, M., Hecht, H. J., Sasse, F., & Reichenbach, H. (1999). N-Oxidation of epothilone A–C and O-acyl rearrangement to C-19- and C-21-substituted epothilones. *Angewandte Chemie International Edition*, 38(13-14), 1971-1974.
- [14] Reichenbach, H., & Höfle, G. (1993). Biologically active secondary metabolites from myxobacteria. *Biotechnology advances*, 11(2), 219-277.
- [15] Reichenbach, H. (2001). Myxobacteria, producers of novel bioactive substances. *Journal of Industrial Microbiology and Biotechnology*, 27(3), 149-156.
- [16] Reichenbach, H., & Dworkin, M. (1981). Introduction to the gliding bacteria. In *The prokaryotes* (pp. 315-327). Springer, Berlin, Heidelberg.
- [17] Shimkets, L., & Woese, C. R. (1992). A phylogenetic analysis of the myxobacteria: basis for their classification. *Proceedings of the National Academy of Sciences*, 89(20), 9459-9463.
- [18] Shimkets, L. J. (1990). Social and developmental biology of the myxobacteria. *Microbiology and Molecular Biology Reviews*, 54(4), 473-501.
- [19] Wireman, J. W., & Dworkin, M. (1975). Morphogenesis and developmental interactions in myxobacteria. *Science*, 189(4202), 516-523.
- [20] Weissman, K. J., & Müller, R. (2010). Myxobacterial secondary metabolites: bioactivities and modes-of-action. *Natural product reports*, 27(9), 1276-1295.
- [21] Wenzel, S. C., & Müller, R. (2009). Myxobacteria—‘microbial factories’ for the production of bioactive secondary metabolites. *Molecular BioSystems*, 5(6), 567-574.
- [22] Weissman, K. J., & Müller, R. (2009). A brief tour of myxobacterial secondary metabolism. *Bioorganic & medicinal chemistry*, 17(6), 2121-2136.
- [23] Weissman, K. J., & Müller, R. (2010). Myxobacterial secondary metabolites: bioactivities and modes-of-action. *Natural product reports*, 27(9), 1276-1295.
- [24] Yan, Z. C., Wang, B., Li, Y. Z., Gong, X., Zhang, H. Q., & Gao, P. J. (2003). Morphologies and phylogenetic classification of cellulolytic myxobacteria. *Systematic and applied microbiology*, 26(1), 104-109.
- [25] Zhang, L., Wang, H., Fang, X., Stackebrandt, E., & Ding, Y. (2003). Improved methods of isolation and purification of myxobacteria and

- development of fruiting body formation of two strains. *Journal of microbiological methods*, 54(1), 21-27.
- [26] Zaburanyi, N., Bunk, B., Maier, J., Overmann, J., & Müller, R. (2016). Genome analysis of the fruiting body-forming myxobacterium *Chondromyces crocatus* reveals high potential for natural product biosynthesis. *Appl. Environ. Microbiol.*, 82(6), 1945-1957.